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Regional variations of organophosphorus flame retardants - Fingerprint of large river basin estuaries/deltas in Europe compared with China

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Abstract

This study reports the occurrence and distribution of organophosphorus flame retardants and plasticizer (OPEs) in sediments of eight large river basin estuaries and deltas across Europe. A robust and sensitive OPE analysis method was developed through the



application of an in-cell clean-up in an accelerated solvent extraction and the use of an GC-MSMS System for instrumental analyses. OPEs were detected in all sediment samples with sum concentrations of up to 181 ng g⁻¹ dw. A fingerprinting method was used to identify river specific pattern to compare river systems. The estuaries and deltas were chosen to have a conglomerate print of the whole river. The results are showing very similar OPE patterns across Europe with minor differences driven by local industrial input. The European estuary concentrations and patterns were compared with OPEs detected in the Xiaoqing River in China, as an example for

a region with other production, usage and legislative regulations. The Chinese fingerprint differed significantly from the overall European pattern.

Introduction

Sediments provide important services in the estuarine ecosystem. They are habitats for a variety of species that form the basis for local food-webs (Kennish, 1992). Due to this importance and their place at the “start” of the food-web, sediments have often been discussed as starting point of bioaccumulation and magnification of lipophilic contaminants (Ernst et al., 1988; Kennish, 1992). Mid- to non-polar compounds adsorb to particles and are deposited on the sea floor (Ernst et al., 1988) where they can be immobilized and stored or enter the food-web. Therefore, sediments can function as sink but also as a secondary source for contaminants (Laane et al., 2013).

Organophosphate esters (OPEs) are widely used as flame retardant and plasticizers in a variety of products such as electronic equipment, furniture, textiles, isolation material and wires (Muir, 1984). Additionally, they are used in up to 15% (by weight) as additives in hydraulic fluids, lubricants and antifoaming agents (Hartmann et al., 2004). With the restriction of polybrominated diphenylethers (PBDEs), organophosphate based flame retardants have become a focus for the polymer industry (van der Veen and Boer, 2012). A result of that is a strong increasing of the production and consumption of OPEs as flame retardants on the global market (van der Veen and Boer, 2012; Wang et al., 2010).

OPEs are primarily used additively in products, which mean they are not chemical bound to the product, which aids the leach-out into the environment (Staaf and Östman, 2005). Risk assessments of several, especially chlorinated, OPEs have shown a potential for carcinogenic effects, acute toxicity as well as high environmental persistence (Reemtsma et al., 2008; Waaijers and Parsons, 2016; Wang et al., 2015). OPEs primarily enter the aquatic environment through atmospheric deposition and leaching from waste water treatment, from where they can be transported into marine areas (Bollmann et al., 2012; Wolschke et al., 2015). Some OPEs travel in

part attached to particles (Wolschke et al., 2016, Sühring et al., 2016) which allows for sinking and accumulating in sediments (Giulivo et al., 2017). Based on their physical-chemical properties, especially the longer chained and the chlorinated OPEs have the potential to accumulate in sediments (Zhang and Sühring et al. 2016). However, the occurrence and distribution of organophosphate flame retardants in sediments is still not sufficiently understood. The presented study focused on the accumulation of OPE sediments from seven major European estuaries. Estuaries were chosen as study areas because they represent a conglomeration over the whole river and allow fingerprinting of the river contamination profile. At the same time estuaries are very specific unique ecosystems which have to handle the fluctuating salinity gradients with moving turbidity and nutrient conditions (Martin and Brun-Cottan, 1988).

In this study, we compared seven European river basins to identify contamination levels and river specific patterns across Europe. Europe has relatively limited production of OPEs and has strict environmental legislations that prevent the production or use of particularly hazardous OPEs (European Commission, 2014). To compare the contamination with a different catchment area and legislative regulation, sediments samples from an estuary in north east China were analyzed.

Materials and methods

Sample collection

29 surface sediment samples were collected from eight large estuaries and delta in Europe. The samples were collected from 2013 to 2015 by stainless steel grab sampler from ship or from shore at low tide. The sediment samples were stored at $-20\text{ }^{\circ}\text{C}$ prior to analysis. Sampling sites are shown in Fig. 1. From the Xiaoqing River in China 5 samples were collected in 2014.

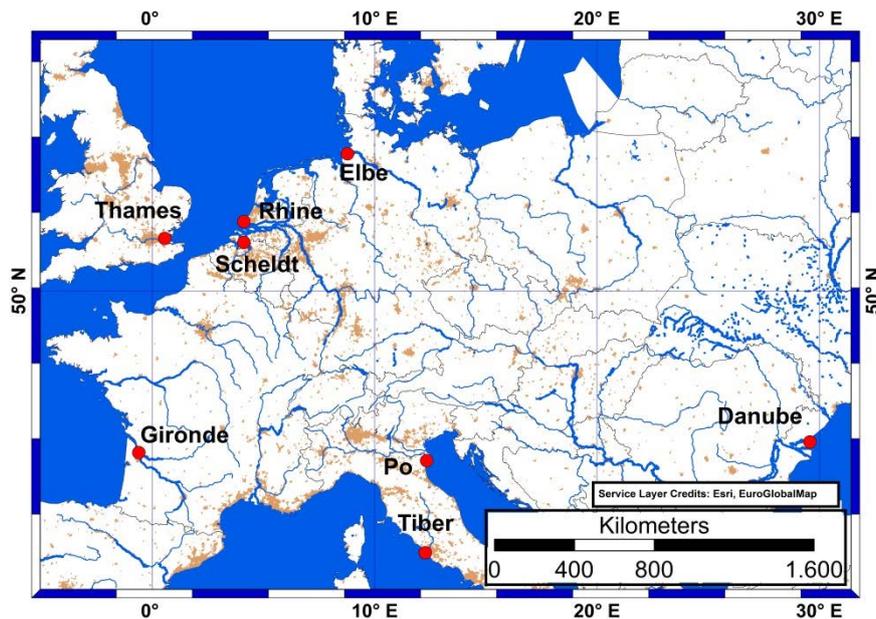


Figure 1: European sampling sites

Sample Preparation

The extraction and clean-up were adapted from a method presented by Sühling et al. (2016). Larger pieces (> 2 mm) were removed by hand from the sediment samples. The samples were homogenized with anhydrous Sodium sulfate (Na_2SO_4) using an agate mortar. The samples were extracted using accelerated solvent extraction (Thermo Fisher ASE-350) with an in-cell clean-up (Sühling et al., 2016). 22 mL stainless steel ASE cells were filled with 7 g 10% deactivated silica gel, 2 g activated Copper and 5-15 g dried sediment. All samples were spiked with mass labelled surrogate standards TCEP-d12, TBP-d27, TPhP-d15.

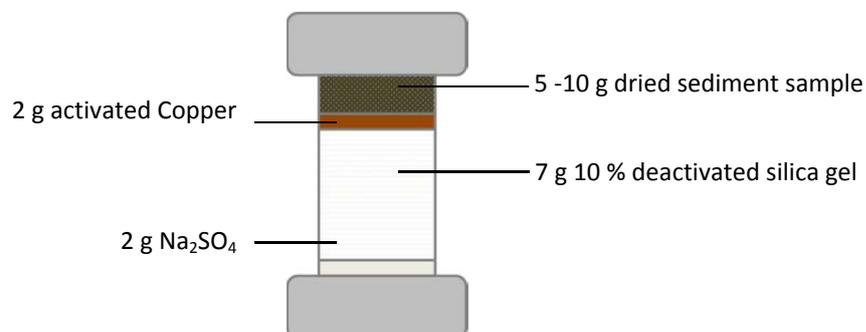


Figure 2: Schematic of the ASE in-cell clean up

The cells were extracted sequentially in two fractions. The first fraction was extraction with following parameters: 2 cycles, hexane as solvent, pressure (nitrogen): 9 bar, temperature: 100 °C, pressure: 120 bar, heat: 5 min, static (2x): 8 min, flush: 100%, purge: 60 sec. The second fraction was extracted using the same conditions with 90:10 Dichloromethane:Acetone as solvent. This second fraction (containing the OPEs), was reduced in volume and subjected to a further clean-up by a 2.5 g 10% water deactivated silica gel column. For elution 20 mL of acetone/DCM (1:1 v/v) were used and the sample reduced to a volume of 150 µL under a gentle stream of heated nitrogen (40°C). Finally, 500 pg (absolute) ¹³C-PCB-141 and ¹³C-PCB-208 were added as injection standards to each sample. Recoveries of deuterated standards, extraction efficiency, blanks and reproducibility were tested with and without matrix, during method validation. All samples were analyzed in duplicates.

Separate sample aliquots were dried to constant weight (at 105 °C) for the gravimetric determination of water content as well as the subsequent analysis of total organic carbon (TOC). TOC was measured using a LECO RC612 multiphase carbon/hydrogen/moisture determinator combustion method at 400 °C.

Instrumental analysis

The samples were analyzed on a GC-MS/MS system (Agilent 7010) in electron impact ionization mode (EI) equipped with two DB-35MS columns (15 m×0.25 mm i.d. × 0.25 µm film thickness, J&W Scientific) connected to a purge unit for backflush after each run. The injector was operated in pulsed-splitless mode (injection pulse 20 psi for 2 min) with an inlet temperature program as follows: 60 °C for 0.1 min and 300 °C min⁻¹ until 300 °C and held for a final 20 min. The GC oven program was as follows: initial 60 °C for 4 min, 25 °C min⁻¹ until 100 °C, 7 °C min⁻¹ until 310 °C and held for 1 min. The backflush was performed at 310 min with a flow of 1.5 ml min⁻¹

at the first column for 5 min. The temperature of the MS transfer line was held at 280 °C. The ion source and quadrupole temperatures were 150 °C.

Quality assurance and quality control (QA/QC)

Because of the widespread presence of OPEs in a variety of laboratory equipment, the use of rubber and plastic materials was avoided to minimize blank contamination during the transport, storage and treatment of the samples. All glassware was cleaned prior to use by a laboratory dishwasher, baked at 250 °C and rinsed with acetone. Na₂SO₄ was cleaned by Soxhlet extraction with DCM for 12 h and baked at 450 °C. Blank samples were analyzed with every batch of 10 samples. Detected blanks were at least one order of magnitude below the measured concentrations for all of the target compounds. Absolute blank values ranged from 0.1±0.02 ng for TiBP to 0.5±0.15 ng for TCPP for the whole method. Method detection limits (MDLs) were derived from either the mean blank values plus three times the standard deviation or at a signal-to-noise ratio of 3 (S/N=3), whichever approach yielded the higher value. Based on a sample amount of 10 grams, MDLs ranged from 37 pg g⁻¹ for TiBP to 250 pg g⁻¹ for TCPP. The average recoveries of the surrogates were 76±18% for TCEP-d12, 56±18% for TnBP-d27, and 74±16% for TPhP-d15. All reported concentrations were corrected for blanks and recoveries.

Results and Discussion

Analytical method

The use of a GC-MS/MS system with a (mid-column) backflush system compared to classic GC-MS systems provided a strong increase of condition stability, while reducing matrix effects. After each run the first half part of the column was flushed backwards with high temperature carrier gas to eliminate matrix debris on the column. These advantages compared with the GC-MS/MS-System lead to a more robust analysis with increased sensitivity and selectivity for the analyses of

OPEs in environmental samples (Wolschke et al., 2016). Due to the wide range of physicochemical properties, instability of compounds and blank contamination, using classic GC-MS, it has been challenging to develop an extraction and cleanup method for complex sediment matrices that reduces the matrix effects enough while not impairing the analytes. With the describe advantage of the GC-MSMS method with mid-column backflush the clean-up process can be simplified.

Environmental occurrence and fate

OPEs were detected in all analyzed sediment samples. The sum concentrations for the European river sediments ranged from 2.5 ng g⁻¹ dry weight (dw) at an up-stream position in the river Gironde to 181 ng g⁻¹ dw in the Belgian river Scheldt. In general, the absolute concentrations were highly variable between the individual rivers. The concentrations are comparable to other studies from Europe. In Greece sediment concentrations in the Evrotas river between 10.5 – 248 ng g⁻¹ dw were reported (Giulivo et al. 2017). In the Adige river, Italy OPE concentrations in sediments ranged from 11.5 – 549 ng g⁻¹ dw and in Slovenia in the Sava river basin from 0.31-310 ng g⁻¹ dw (Giulivo et al. 2017). Interestingly, Brandsma et al. (2015) reported lower OPE concentrations in the river Scheldt river (<0.1 – 19.6 ng g⁻¹ dw) compared to this study this study. A possible explanation could be that the OPE concentrations have increased over the years, as the samples from Brandsma et al. (2015) were collected in 2008, compared to 2014 for this study. But local differences in accumulation capacities and discharge patterns could also be a reason for the differences in detected concentrations. To ensure comparability between the sampling sites, the contamination levels and patterns in this study are therefore being discussed normalized for TOC, (Figure 3).

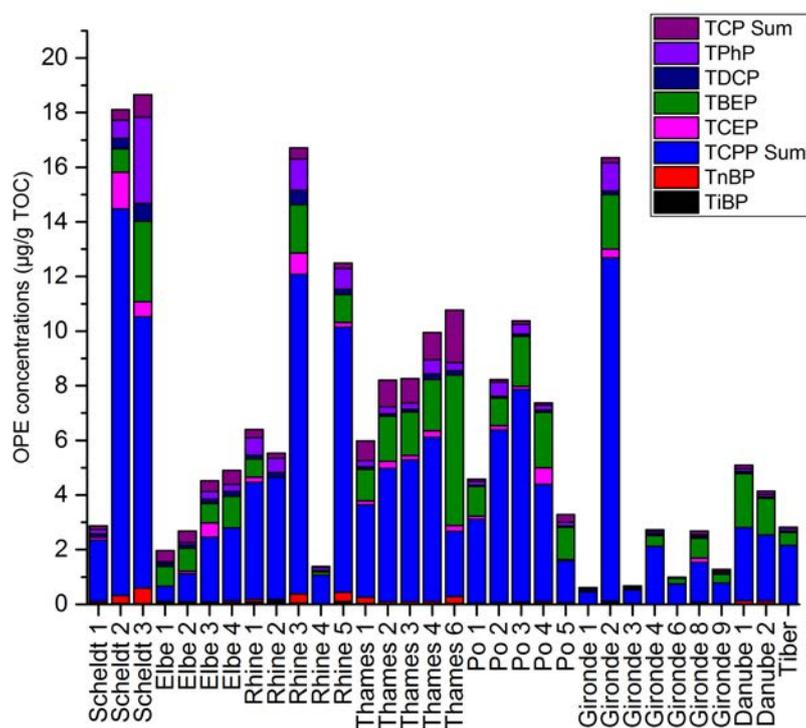


Figure 1: Individual sediment concentrations along the estuaries in ng g^{-1} TOC dry weight

The results indicated that the river Scheldt with OPEs concentration up to $18 \mu\text{g g}^{-1}$ TOC is the most contaminated river for OPEs in Europe (Figure 3), followed by the Rhine, Thames and Po. The rivers Elbe, Danube, Tiber and Gironde displayed overall lower OPEs contaminations (Figure 3). These differences in concentrations was expected, the catchment area of the rivers Scheldt (Antwerp), Rhine (Rotterdam, Ruhr area) and Thames (London) are highly populated and strongly influenced by industry. An interesting exemption to this observation were the high OPEs concentrations ($17 \mu\text{g g}^{-1}$ TOC) at a sampling station in the river Gironde near the city of Blaye (Gironde 2 in Figure 3), suggesting that there might be local sources for OPEs in that area. In all samples, the chlorinated tris(2-chloroethyl) phosphate (TCPP) was the dominating (highest concentrated and detection frequency) compound with a contribution to the sum of OPEs up to 70% (average 63%). The high contributions of TCPP were congruent with findings reported in other matrices from Europe (Malarvannan et al., 2015; Möller et al., 2011; Regnery and Püttmann, 2009; Wolschke et al., 2016; Wolschke et al., 2015). The second highest concentrated and prevalent compound was Tris(2-butoxyethyl) phosphate (TBEP) with an average

contribution of 18%. However, compared to TCPP, TBEP showed a higher variability of contributions to sum OPEs in different rivers. In Elbe and Gironde TBEP had the highest contributions to the OPEs contamination pattern with up to 30%, whereas its contributions in the river Scheldt were lower (0-15%). The reason is not clear, and it can indicate a higher input of TBEP in this river catchments but a specific usage or production is not known in this area. Another OPE with river specific contamination pattern was Tri-cresly-phosphate (TCP) - in the rivers Elbe and Thames its contribution was comparably high with around 10 % compared to the average contribution in other rivers of only 3%. These findings indicated that for many OPEs, the contamination patterns are location specific, forming an individual fingerprint.

To compare the contamination patterns from a use area (Europe) and a production area (China), five samples from the Xiaoqing River, Shandong, China were analysed. The catchment area of the river encompasses industrial areas around the cities Weifang, Zibo and Jinan.

The OPE patterns in the Xiaoqing were noticeably different compared to the patterns found in European rivers (Figure 4). The major component was TCEP (46 %), a component that is restricted in Europe (European Commission, 2014). Consequently, the contributions of TCEP in Europe are significantly lower ($p < 0.01$). At the same time, the major component in Europe, TCPP, only contributed around 20 % to the OPE pattern in China (Figure 4). TCPP is the main replacement for TCEP (World Health Organization, 1998). The differences in patterns between Europe and China highlight the effectiveness of restrictions of TCEP, but also the problem of “regrettable substitution”, since the overall use of OPEs has not declined but the pattern has merely shifted to other OPE components. An interesting similarity between the OPE patterns in China and Europe were the high contribution of TCP which were also present in Elbe and Thames.

Fingerprint analyses

To analyze individual contamination “fingerprints” of the different estuaries, a method for the “Fingerprint Analysis of Contaminant Data” (Russell H. Plumb, 2004) by the United States Environmental Protection Agency (EPA) was used. The average concentrations [$\text{pg g}^{-1} \text{ dw}$] of each river were used to determining the contribution to the sum contamination in the investigation area (i.e. specific estuary):

$$con_{xi}[\%] = \frac{c_{xi}[\text{pg g}^{-1} \text{ dw}]}{\sum_{i=1}^n c_{xi}[\text{pg g}^{-1} \text{ dw}]}$$

With

$con_x[\%]$: Contribution in % of compound $X_{i=1-n}$ to the sum concentration in the investigation area

$c_{xi}[\text{pg g}^{-1} \text{ dw}]$: Concentration of compound $X_{i=1-n}$ in $\text{pg g}^{-1} \text{ dw}$

$d \sum_{x=1}^n c_x[\text{pg g}^{-1} \text{ dw}]$: Sum concentration of all compounds X in the investigation area in $\text{pg g}^{-1} \text{ dw}$

The resulting contribution of individual substances to the contamination pattern in an investigated area was then used to create a “fingerprint” of the contamination patterns in that area (Figure 4). For comparison of the different investigation areas, the determined patterns were correlated with each other, using a Pearson correlation analyses (Table 1). As expected, the OPE patterns in European rivers were very similar with Pearson correlation coefficients between 0.86 and 0.99 (Table 1, Figure 2). Particularly, the Scheldt and Rhine with a similar catchment and in parts water mixing were highly correlated ($r > 0.99$); but high correlations were also found for the Gironde in France and the Po in Italy ($r > 0.99$). The other “group” was the Elbe estuary in Germany, the Thames in England and the Danube Rumania. These had highly intercorrelated patterns ($r > 0.99$), but were slightly less correlated with the Scheldt, Rhine, Gironde and Po patterns ($r < 0.97$) (Table 1).

The pattern in China was significantly different from all analysed European estuaries ($p < 0.01$) with correlation coefficients between 0.08 (Danube) and 0.27 (Scheldt) (Table 1). The different profile highlighted the differences between production and usage areas, as well as the differences

in legislations. The presented study showed that a common market and harmonised regulations generates a comparable fingerprint of contamination and can effectively reduce contamination from specific hazardous compounds such as the chlorinated TCEP in a large area.

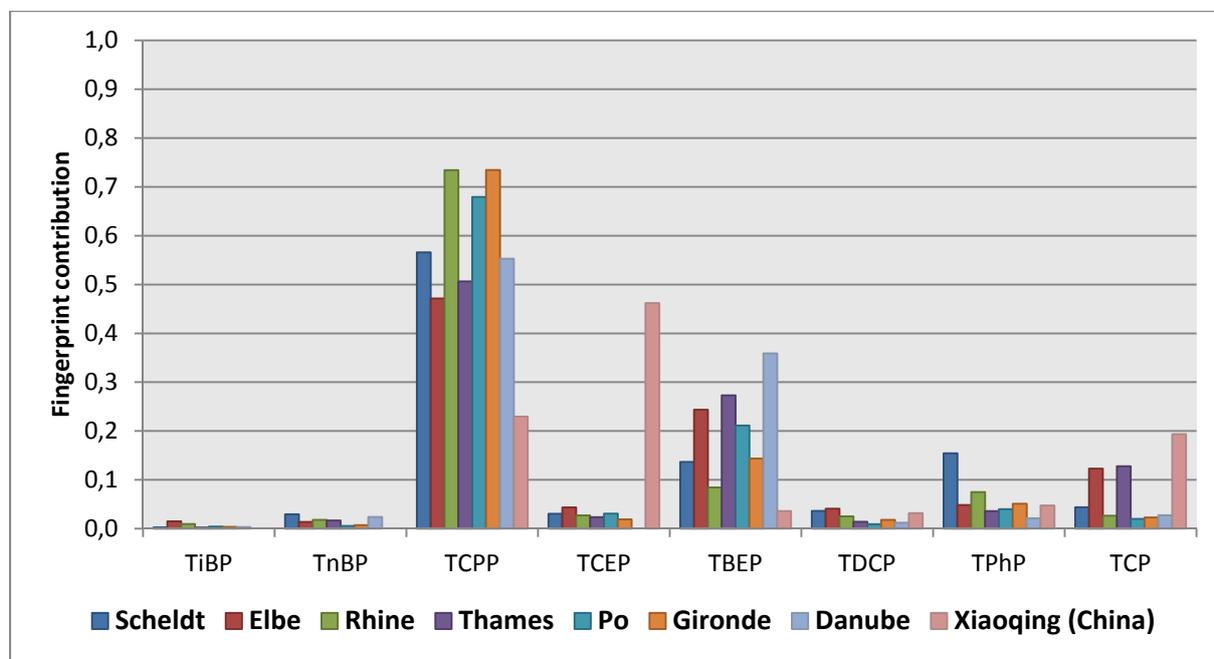


Figure 2: Fingerprint of different estuaries/deltas

| | | Elbe | Rhine | Thames | Po | Gironde | Danube | Xiaoqing (China) |
|----------------|--------------|----------------|----------------|----------------|----------------|----------------|----------------|------------------|
| Scheldt | Pearson Korr | 0.91063 | 0.99836 | 0.89453 | 0.97627 | 0.99365 | 0.86159 | 0.26592 |
| | Sig. | 1.67E-03 | 1.09E-08 | 2.71E-03 | 3.28E-05 | 6.38E-07 | 0.00596 | 0.52441 |
| Elbe | Pearson Korr | 1 | 0.90855 | 0.99786 | 0.96699 | 0.94154 | 0.96978 | 0.21812 |
| | Sig. | | 1.78E-03 | 2.44E-08 | 8.77E-05 | 4.78E-04 | 6.74E-05 | 0.60382 |
| Rhine | Pearson Korr | | 1 | 0.89277 | 0.97602 | 0.99508 | 0.85962 | 0.24445 |
| | Sig. | | | 2.84E-03 | 3.39E-05 | 2.97E-07 | 0.00621 | 0.55959 |
| Thames | Pearson Korr | | | 1 | 0.9584 | 0.92891 | 0.97469 | 0.19115 |
| | Sig. | | | | 1.74E-04 | 8.51E-04 | 3.98E-05 | 0.65023 |
| Po | Pearson Korr | | | | 1 | 0.99254 | 0.94825 | 0.22129 |
| | Sig. | | | | | 1.03E-06 | 3.33E-04 | 0.59843 |
| Gironde | Pearson Korr | | | | | 1 | 0.90517 | 0.22913 |
| | Sig. | | | | | | 0.00198 | 0.58519 |
| Danube | Pearson Korr | | | | | | 1 | 0.0898 |
| | Sig. | | | | | | | 0.83253 |

Table 1: Pearson correlations of fingerprint analyses

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